

A Statistical Analysis of Chronic Liver Disease Diagnosis with Noninvasive Biomarkers

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Keywords: Biomarker, HCV, Fibrosis, Cirrhosis, Multinomial Logistics Regression, LASSO, Ridge Regression.


Abstract: Chronic hepatitis C virus infection (CHC) can cause life-threatening liver diseases such as cirrhosis and fibrosis. This study aims to investigate how noninvasive serum biomarkers can aid in CHC infected liver disease diagnosis. Previous studies have researched various combinations of serum biomarkers. This study examines the diagnosing effect of a different combination of serum biomarkers on CHC patients. A multinomial logistics regression model is employed to make a secondary analysis of the HCV dataset. We use LASSO, stepwise regression, and ridge regression for model selection. Average accuracy, sensitivity, precision, and specificity are calculated to evaluate model performance. Our statistical analysis resulted in high accuracy and specificity. The average accuracy and sensitivity for predicting cirrhosis have both achieved 99%. The average specificity for predicting fibrosis has attained 95%. Our statistical analysis result implicates that future research on CHC diagnosis can analyze different combinations of serum biomarkers or even genetic markers.

1 INTRODUCTION

The hepatitis C virus (HCV) was discovered by Nobel Prize winning researchers Harvey J. Alter, Michael Houghton, and Charles M. Rice (Masucci, Hedestam 2020). Alter, Houghton, and Rice also determined HCV to be caused by an RNA virus from the Flavivirus family (Masucci, Hedestam 2020). HCV can cause both long-term and short-term liver disease, but more than half of the infected patients will suffer from chronic infection of HCV (Centers for Disease Control and Prevention 2020). According to American Centers for Disease Control and Prevention (CDC), chronic hepatitis C can pose severe and life-threatening health problems like fibrosis and cirrhosis (Centers for Disease Control and Prevention 2020). Liver fibrosis is caused by wounded tissue healing in response to HCV inflicted damage and is characterized by the excessive accumulation of extracellular matrix (ECM) proteins (Khatun and Ray 2019). HCV is an infectious virus and there are currently no vaccines for hepatitis C virus (Centers for Disease Control and Prevention 2020). Thus, it is imperative to investigate accurate and specific

biomarker predictors of CHC infection stage. Although liver biopsy is considered the gold standard for diagnosing CHC infected liver disease, some research focused on alternative diagnosing methods using noninvasive serum biomarkers (Pár, Vincze and Pár 2015, Sebastiani, Gkouvatzos and Pantopoulos 2014, Shahid, Idrees, Nasir, Raja, Raza, Amin, Rasul and Tayyab 2014, Valva, Rios, Matteo and Preciado 2016). With more patient-friendly and noninvasive avenues to diagnose CHC infection stage, effective treatment can be applied to cure patients quickly (Centers for Disease Control and Prevention 2020) without excessive pain.

Previous studies applied various statistical models and machine learning methods to research diagnosing effect of serum biomarkers (López, Manzano, et al. 2020, Fornas, Ampurdanès, Llovet, Aponte, Quintó, et al. 2002, Hoffmann, Bietenbeck, Lichtinghagen and Frank Klawonn 2018, Peschel, Grimm, Gülow, Müller, Buechler and Weigand 2020, Staufer, Dengler, et al. 2017, Syafa'ah, Zulfatman, Pakaya and Lestandy 2021, Valva, Casciato, Carrasco, Gadano, et al. 2011). An early study published in 2002 utilized a multiple logistics regression model to predict CHC

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infected liver fibrosis (Forns, Ampurdanès, Llovet, Aponte, Quintó, et al. 2002,). This study has identified age, gamma-glutamyl transpeptidase (GGT), cholesterol, platelet count, and prothrombin time as important predictors of fibrosis (Forns, Ampurdanès, Llovet, Aponte, Quintó, et al. 2002,). However, the predictive accuracy was relatively low (0.66) in the validation set, and the study did not include p-values in the final multivariate model (Forns, Ampurdanès, Llovet, Aponte, Quintó, et al. 2002,). Other biomarkers like albumin (ALB) (Staufér, Dengler, et al. 2017), chemerin (CHE) (Peschel, Grimm, Gülow, Müller, Buechler and Weigand 2020), HA, PIIINP, and TGF-β1 (Valva, Casciato, Carrasco, Gadano, et al. 2011) have all been identified as relatively accurate predictors of CHC infected liver disease stage. Effective CHC liver disease stage predicting indexes such as AST to platelet ratio index (APRI), WFA-M2BP, and ELF score have also been studied individually or together as covariates (Fujita, Kuroda, Morishita, Oura, Tadokoro, Nomura, Yoneyama, et al. 2018, Wai, Greenson, Fontana, Kalbfleisch, Marrero, Conjeevaram, and Lok 2003). Some studies explored machine learning methods other than logistics regression (Hoffmann, Bietenbeck, Lichtinghagen and Frank Klawonn 2018, Syafa'ah, Zulfatman, Pakaya and Lestandy 2021). The original paper (Hoffmann, Bietenbeck, Lichtinghagen and Frank Klawonn 2018) used ctree and rpart algorithm on a subset of the HCV dataset biomarkers (Lichtinghagen, Klawonn and Hoffmann), but the highest accuracy did not exceed 80%. Another paper used naïve Bayes classifier, neural network, and random forest (Syafa'ah, Zulfatman, Pakaya and Lestandy 2021) to model all biomarkers in the HCV dataset (Lichtinghagen, Klawonn and Hoffmann). The predictive accuracy using neural network achieved as high as 95.12% (Syafa'ah, Zulfatman, Pakaya and Lestandy 2021). Our study uses

multinomial logistics regression to analyze the HCV dataset (Lichtinghagen, Klawonn and Hoffmann) and evaluates its performance on predicting CHC infected liver disease stage. The paper is organized in the following order: introduction to our data source, elucidation of research variables, explanation of the statistical method, statistical analysis result, limitations, and conclusions.

2 DATA SOURCE

The dataset used in this paper is obtained from UCI Machine Learning Repository, a free machine learning database established in 2019 (Dua and Graff). UCI Machine Learning Repository (Dua and Graff) offers many high-quality datasets that can be used for academic research. The HCV dataset contains 615 samples and 14 variables: CHC infection stage (Category), age, sex, albumin level (ALB), alkaline phosphatase level (ALP), alanine aminotransferase level (ALT), aspartate aminotransferase level (AST), bilirubin level (BIL), serum cholinesterase level (CHE), cholesterol level (CHOL), creatinine level (CREA), gamma-glutamyl transferase level (GGT), and overall protein level (PROT). This HCV dataset was originally used in a study by Hoffmann, Bietenbeck, Lichtinghagen, and Klawonn (Hoffmann, Bietenbeck, Lichtinghagen and Frank Klawonn 2018).

3 RESEARCH VARIABLES

The response variable is CHC infection stage (Category), a categorical variable indicating diagnosis result of CHC infected liver disease. The 12 covariates used in this study are listed in Table 1.

Table 1: Variables Explained.

Variable	Meaning	Type	Range
Category	CHC infected liver disease stage	Categorical	0=Blood Donors, 1=Hepatitis, 2=Fibrosis, 3=Cirrhosis
Sex	Gender		Female, Male
Age	Samples' Age		[23.0, 77.0]
ALB	Albumin level		[23.0, 82.2]
ALP	Alkaline phosphatase level		[11.3, 416.6]
ALT	Alanine aminotransferase level		[0.9, 118.1]
AST	Aspartate aminotransferase level		[12.0, 324.0]
BIL	Bilirubin level		[1.8, 209.0]

CHE	Serum cholinesterase level	Numerical	[1.42, 16.41]
CHOL	Cholesterol level		[1.43, 9.67]
CREA	Creatinine level		[8.0, 1079.1]
GGT	Gamma-glutamyl transferase level		[4.5, 650.9]
PROT	Overall protein level		[51.0, 86.5]

4 STATISTICAL METHOD



Figure 1: Implementation of statistical analysis.

First, we processed our dataset before statistical analysis. The dataset initially consists of 615 observations. We removed the NAs and the patient ID indicator column. There are 589 observations left after removing the NAs. There are originally five categories in the response variable: 0=Blood Donor, 0s=suspect Blood Donor, 1=Hepatitis, 2=Fibrosis, and 3=Cirrhosis. The category “0=Blood Donor” means healthy samples that are not diagnosed with CHC infected liver disease. The category “0s=suspect Blood Donor” indicates it is undetermined whether the sample contracted CHC infected liver disease or not. The other three categories represent different diagnosis stages of CHC infected liver disease. “1=Hepatitis” is the least severe of the three categories, and “3=Cirrhosis” is the most severe. We removed observations classified as “0s=suspect Blood Donor” because our research interest focuses on predicting diseased versus healthy samples.

After processing the data, we performed exploratory data analysis. The dataset was initially very imbalanced. About 90.3% of the samples are classified as “0=Blood Donor” (non-diseased samples). About 3.4% of the samples are classified as “1=Hepatitis”. About 2.1% of the samples are classified as “2=Fibrosis”. About 4.1% of the samples are classified as “3=Cirrhosis”. To balance the data, we randomly replicated examples with replacement so that each category (“0=Blood Donor”, “1=Hepatitis”, “2=Fibrosis”, “3=Cirrhosis”) in the response variable contains the same number of

samples. After we balanced the data, there are 1432 samples in total and 526 samples for each of the four categories in the response variable, CHC infected liver disease stage. We computed the range for all numerical variables and summarized them in Table 1.

A multinomial logistics regression model was fit using variables listed in Table 1. Some previous studies also used the multinomial logistics regression model, but they used different combinations of biomarkers or liver disease indexes other than this study. First, the data was modeled using the entire balanced dataset without training. The corresponding regression coefficients and p-values were calculated to exclude non-significant variables from the model. We used a significance level of $\alpha = 0.05$. After that, we performed forward, backward and bidirectional model selection to fine-tune our model. LASSO and cross-validation were implemented to eliminate unimportant variables. We fitted a multinomial logistics regression model with all selected variables. We randomly partitioned the samples into training and testing sets 100 times to estimate the average model performance. We computed mean accuracy, mean recall sensitivity, mean precision, and mean specificity. The corresponding standard deviations of the model accuracy, recall sensitivity, precision, and specificity are also calculated after 100 iterations. In addition, we also performed multinomial logistics regression with L2 penalty (ridge regression). The dataset was again randomly split into training versus validation sets 100 times. Corresponding statistics for ridge regression performance are also computed. Mean accuracy, recall sensitivity, precision, and specificity are calculated. The standard deviations of model accuracy, recall sensitivity, precision, and specificity are computed. All data analysis in this study was done using software R version 4.1.0 (<https://cran.r-project.org/>).

5 RESULT

Multinomial logistics regression was employed to make a statistical analysis of the relationship between age, sex, albumin level (ALB), alkaline phosphatase

level (ALP), alanine aminotransferase level (ALT), aspartate aminotransferase level (AST), bilirubin level (BIL), serum cholinesterase level (CHE), cholesterol level (CHOL), creatinine level (CREA), gamma-glutamyl transferase level (GGT), overall protein level (PROT) and the response variable CHC infected liver disease stage. It can be found that if all other predictor variables are held constant, the odds of “1=Hepatitis” occurring decreased by 1.05 (95% CI [-1.28, -0.824]) for a one-unit increase in ALP. The odds of “2=Fibrosis” occurring decreased by 0.994 (95% CI [-1.22, -0.768]) for a one-unit increase in ALP. The odds of “3=Cirrhosis” occurring decreased by 0.349 (95% CI [-0.48, -0.218]) for a one-unit increase in ALP. It was found that if all other predictor variables are held constant, the odds of “1=Hepatitis” occurring increased by 0.218 (95% CI [0.172, 0.265]) for a one-unit increase in AST. The odds of “2=Fibrosis” occurring increased by 0.236 (95% CI [0.189, 0.283]) for a one-unit increase in AST. The odds of “3=Cirrhosis” occurring increased by 0.201 (95% CI [0.143, 0.259]) for a one-unit increase in AST. It was also found that if all other predictor variables are held constant, the odds of “1=Hepatitis” occurring increased by 0.747 (95% CI [0.502, 0.993]) for a one-unit increase in BIL. The odds of “2=Fibrosis” occurring increased by 0.669 (95% CI [0.424, 0.913]) for a one-unit increase in BIL. The odds of “3=Cirrhosis” occurring increased by 0.449 (95% CI [0.137, 0.76]) for a one-unit increase in BIL. It was also found that if all other predictor variables are held constant, the odds of “1=Hepatitis” occurring increased by 2.43 (95% CI [0.317, 4.55]) for a one-unit increase in CHE. The odds of “2=Fibrosis” occurring increased by 2.31 (95% CI [0.193, 4.42]) for a one-unit increase in CHE. The odds of “3=Cirrhosis” occurring decreased by 7.95 (95% CI [-10.6, -5.26]) for a one-unit increase in CHE. It was shown that, if all other predictor variables are held constant, the odds of “1=Hepatitis” occurring decreased by 4.52 (95% CI [-6.72, -2.32]) for a one-unit increase in CHOL. The odds of “2=Fibrosis” occurring decreased by 4.91 (95% CI [-7.11, -2.72]) for a one-unit increase in CHOL. The odds of “3=Cirrhosis” occurring decreased by 2.66 (95% CI [-4.33, -0.99]) for a one-unit increase in CHOL.

Table 2 summarizes our model performance calculated using fully balanced data before we train the model. The accuracy and specificity of the model are consistently high across all four categories.

Table 5 and 6 (see the following pages) present the regression coefficients of selected variables and their corresponding p-values. Variables of interest,

ALB, ALP, AST, BIL, CHE, CHOL, GGT, and PROT, are significant ($p < 0.05$) for all 3 stages of CHC infected liver disease. Variable CREA is only significant for “3=Cirrhosis”. Variable ALT is only significant for “1=Hepatitis” and “2=Fibrosis”.

Table 2. Model performance using full data.

	Accurac y	Sensitivit y	Precisio n	Specificit y
0=Blood Donor	0.998	1.000	0.992	0.998
1=Hepatitis	0.915	0.847	0.808	0.937
2=Fibrosis	0.915	0.814	0.856	0.951
3=Cirrhosis	0.999	0.998	1.000	1.000

We implemented backward stepwise model selection, forward stepwise model selection, and bidirectional model selection. No variables were eliminated. The cross-validation result from LASSO did not indicate a single variable to be unimportant for all four categories of the response variable, but LASSO ruled out variable CREA for “0=Blood Donors” and “1=Hepatitis”. No variables were eliminated.

We randomly partitioned the data 100 times into training and validation sets to improve model performance. Table 3 lists the mean and standard deviation of our final model performance after training. The standard deviations are consistently small for all entries. The average model performance statistics are consistently high across all four categories of CHC infected liver disease stage. However, the average accuracy, sensitivity, precision, and specificity computed for the testing set are not significantly different from the performance of the initial untrained model.

Table 3. Model performance in the testing set (no L2 penalty).

	Accuracy Mean (SD)	Sensitivity Mean (SD)	Precision Mean (SD)	Specificity Mean (SD)
0=Blood Donor	0.996 (0.00317)	1.00 (0)	0.984 (0.0127)	0.995 (0.00420)
1=Hepatitis	0.915 (0.0129)	0.844 (0.030)	0.807 (0.0429)	0.937 (0.0145)
2=Fibrosis	0.914 (0.0127)	0.813 (0.0382)	0.856 (0.0283)	0.951 (0.0103)
3=Cirrhosis	0.999 (0.00213)	0.994 (0.00837)	1.00 (0)	1.00 (0)

Finally, we added an L2 penalty to fit a ridge multinomial logistics regression. The average ridge model performance is summarized in Table 4 below. The standard deviations of the accuracy, sensitivity, precision, and specificity after 100 iterations are very

similar to those calculated without adding an L2 penalty. However, it seems that model precision and sensitivity have drastically declined for predicting “1=Hepatitis” and “2=Fibrosis”, decreasing from over 90% to less than 70%. We also observed a slight decline in model accuracy and specificity across all four categories in the response variable. The multinomial logistics regression model without L2 penalty has higher accuracy and specificity in terms of model performance. Nevertheless, adding an L2 penalty may address the potential problem of overfitting. Since we randomly replicated samples to account for the imbalanced sample distribution in the response variable, there might be potential issue of overfitting.

Table 4. Model performance in the testing set (with L2 penalty).

	Accuracy	Sensitivity	Precision	Specificity
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)
0=Blood Donor	0.975 (0.0179)	0.927 (0.0558)	0.981 (0.0105)	0.993 (0.0036)
1=Hepatitis	0.826 (0.02)	0.655 (0.0453)	0.64 (0.0509)	0.881 (0.0179)
2=Fibrosis	0.839 (0.0198)	0.691 (0.0408)	0.646 (0.0648)	0.885 (0.0217)
3=Cirrhosis	0.975 (0.00621)	0.939 (0.0178)	0.965 (0.0148)	0.988 (0.00513)

Table 5: Coefficients and p-values of hepatitis and fibrosis.

Variable	Hepatitis		Fibrosis	
	Coefficient	p-value	Coefficient	p-value
Age	-0.013	0.82	0.11	0.058
Sex (male)	-6.4	$< 2.2 \times 10^{-16}$	-6.0	$< 2.2 \times 10^{-16}$
ALB	0.73	0.029	0.72	0.031
ALP	-1.1	$< 2.2 \times 10^{-16}$	-0.99	$< 2.2 \times 10^{-16}$
ALT	-0.56	1.3×10^{-13}	-0.56	1.3×10^{-13}
AST	0.22	$< 2.2 \times 10^{-16}$	0.24	$< 2.2 \times 10^{-16}$
BIL	0.75	2.5×10^{-9}	0.67	8.2×10^{-8}
CHE	2.4	0.024	2.3	0.032
CHOL	-4.5	5.7×10^{-5}	-4.9	1.2×10^{-5}
CREA	-0.059	0.41	-0.099	0.17
GGT	0.30	6.4×10^{-15}	0.28	3.0×10^{-13}
PROT	1.2	4.2×10^{-9}	1.2	5.6×10^{-9}

Table 6: Coefficients and p-values of cirrhosis.

Cirrhosis		
Variable	Coefficient	p-value
Age	0.54	0.056
Sex (male)	-8.2	$< 2.2 \times 10^{-16}$
ALB	-1.6	0.012
ALP	-0.35	1.7×10^{-7}
ALT	-1.1	0.051
AST	0.20	1.4×10^{-11}
BIL	0.45	4.7×10^{-3}
CHE	-7.9	7.1×10^{-9}
CHOL	-2.7	1.8×10^{-3}
CREA	0.13	1.4×10^{-4}
GGT	0.18	3.0×10^{-6}
PROT	1.9	7.5×10^{-7}

6 DISCUSSIONS

Our study supports the hypothesis that the biomarkers listed in Table 1 are significantly associated with CHC infected liver disease stages. Our multinomial logistics regression model included 10 biomarkers to predict CHC infected liver disease stage. The statistical analysis result is highly accurate, sensitive, precise, and specific (see Table 4). Future research can test such combinations for more accurate and specific diagnosing of CHC infected liver disease stages.

However, our study has the following limitations. First, it is conducted from the point of view of statistical analysis. Hence, it requires further reviews from professionals in medical fields, especially clinical fields. Second, the dataset used in our study includes relatively limited patient characteristics. The dataset only has information on patients’ age and gender. Other key biochemical markers for detecting CHC infected liver disease may also be lacking in this dataset. Third, despite the good predictive performance of our final multinomial logistics model, there might be potential overfitting issues. We randomly replicated samples with replacement to ensure that all four categories in the response variable

(CHC infected liver disease stage) contain the same number of samples. Other limitations may include further investigations of pesky points (e.g., outliers, high leverage points) and collinearity issues. Although there do not seem to be many collinearity issues between covariates, it is worth noting that three pairs of covariates have a high Pearson correlation. Specifically, the Pearson correlation is 0.69 between CHE and ALB, 0.63 between GGT and ALP, and -0.54 for CHE and BIL.

7 CONCLUSIONS

In summary, this paper researched how noninvasive serum biomarkers can improve the diagnosis of chronic hepatitis C virus infected liver disease. We addressed the research question by fitting an accurate and specific multinomial logistics regression on the HCV dataset. With enhanced diagnosis efficiency, the effect of treatment could be significantly augmented, and more lives could be saved.

Future research can explore the diagnosis effect of other combinations of non-invasive serum biomarkers. Besides, future research can also investigate the influence of genetic factors in diagnosing CHC infected liver disease. Furthermore, key clinical features other than age and gender can also be incorporated as covariates in the statistical analysis so that more comprehensive clinical applications could be developed.

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