# PROTEIN SECONDARY STRUCTURE PREDICTION USING KNOWLEDGE-BASED POTENTIALS

Saras Saraswathi, Robert L. Jernigan, Andrzej Kloczkowski

Department of Biochemistry, Biophysics and Molecular Biology L.H. Baker Center for Bioinformatics and Biological Statistics 112 Office and Laboratory Building, Ames, IA, 50011, U.S.A.

#### Andrzej Kolinski

Laboratory of Theory of Biopolymers, Faculty of Chemistry, Warsaw University, Pasteura 1, 02-093 Warsaw, Poland

Keywords: Protein secondary structure prediction, Neural networks, Extreme learning machine, Particle swarm optimization.

Abstract: A novel method is proposed for predicting protein secondary structure using data derived from knowledge based potentials and Neural Networks. Potential energies for amino acid sequences in proteins are calculated using protein structures. An Extreme Learning Machine classifier (ELM-PSO) is used to model and predict protein secondary structures. Classifier performance is maximized using the Particle Swarm Optimization algorithm. Preliminary results show improved results.

# **1 INTRODUCTION**

Large scale advances in genome sequencing and resultant availability of large numbers of proteins sequences has given protein secondary structure prediction increasing importance in computational biology. Improvements in secondary structure prediction can lead to progress in protein engineering and drug design. Existing crystallographic techniques are too expensive and time consuming for large-scale determination of protein three-dimensional structures. Prediction of secondary structures might be a useful intermediate step to speed up structure prediction (Lomize, Pogozheva and Mosberg, 1999 and Ortiz, Kolinski, Rotkiewicz, Ilkowski and J. Skolnick, 1999). Secondary structure prediction can assist in gene function and sequence annotation, as well as identification and classification of structures and functional motifs and in identifying malfunctioning structures which cause human diseases.

Several computational methods have been successfully used in secondary structure prediction, of which empirical and machine learning methods have proved to be the most successful. Chou and Fasman (1974), Qian and Sejnowski (1988), Ward, McGuffin, Buxton, and Jones (2003) were followed by numerous others. The GOR method based on information theory was used by Garnier. Osguthorpe, and B. Robson, (1978) and later by Garnier, Gibrat, and Robson (1996). Kloczkowski, K.L. Ting, R.L. Jernigan, and J. Garnier (2002) used evolutionary information in GOR V for improved structure prediction. PredictProtein server (Rost, G. Yachdav, and J. Liu, 2004) uses multiple sequence alignment based neural networks. The PSIPRED algorithm developed by Jones (1999) uses PSI-BLAST (Altschul, T.L. Madden, A.A. Schäffer, J. Zhang, Z. Zhang, W. Miller, and D.J. Lipman, 1997) and neural networks. The Jpred prediction server (Cole, Barber, and Barton, 2008), runs on the Jnet algorithm (Cuff and Barton, 2000). Montgomerie, Sundaraj, Gallin, and Wishart (2006) and Pollastri, Martin, Mooney and Vullo, (2007) developed large scale secondary structure prediction methods using existing structural information and computational methods to claim an accuracy of 90% for sequences with over 30% sequence homology. Kihara (2005) suggested that long-range interactions are an important factor to be considered in order to achieve higher classification accuracy.

Saraswathi S., Jernigan R., Kloczkowski A. and Kolinski A..

DOI: 10.5220/0003086903700375
In Proceedings of the International Conference on Fuzzy Computation and 2nd International Conference on Neural Computation (ICNC-2010), pages 370-375
ISBN: 978-989-8425-32-4
Copyright © 2010 SCITEPRESS (Science and Technology Publications, Lda.)

<sup>370</sup> PROTEIN SECONDARY STRUCTURE PREDICTION USING KNOWLEDGE-BASED POTENTIALS.

We propose a novel strategy for secondary structure prediction using knowledge based potential profiles. A two stage Extreme Learning Machine (ELM) (Huang, Q.Y. Zhu, and C.K. Siew, 2006) classifier called the ELM-PSO is used for classification of secondary structures. Performance is improved using Particle Swarm Optimization (PSO) (Clerc, J. kennedy, 2002).

This paper is organized as follows: Section 2 gives a brief description of the data. Section 3 describes the two-stage ELM-PSO classification technique. Section 4 discusses the results and gives a comparative study followed by conclusions in Section 5.

# 2 DATA GENERATION USING POTENTIAL ENERGY

The dictionary of secondary structure assignment Database of Secondary Structure in Proteins (DSSP) (Kabsch and Sander, 1983) has 8 classes of protein secondary structures. We use only a reduced set of three secondary structures, namely, alpha-helix (H), beta-strand (E) and coil (C). Data is derived based on CABS force-fields (Kolinski, 2004 – algorithm for data generation has been submitted for publication), which includes information pertaining to long and short range interactions between amino acids in proteins. A profile matrix was created using the 513 non-homologous (target) protein sequences from the CB513 data set (Cuff and Barton, 2000), where the sequence homology is less than 30%.

# 3 METHODS AND OPTIMIZATION

In a neural net framework, the input consists of a set of patterns (residues), each having a set of 27 features (profile values), which are normalized to values between 0 and 1. The output consists of three units which correspond to one of three secondary structure elements, represented as a 1 for the class of interest and a -1 for the other two classes. A given input is combined with a bias and a set of weights and is processed through an activation function at the hidden layer level. The output of the hidden layer is combined with another set of weights to yield three outputs. The predicted class is considered as the output which has the maximum value, which corresponds to choosing the output with the smallest mean-squared error. An Extreme Learning Machine (ELM) (Huang, Zhu, and Siew, 2006) classifier, which is a form of a Neural Network, is used for classification. PSO is used to tune the parameters of the ELM. The data was also evaluated using Support Vector Machine (SVM) and Naïve Bayes (NB) algorithms using the WEKA (Witten and Frank,2005) software tool for classification.

The profile data consists of 27 features for each of N amino acids, where N is the number of residues in a single protein. Of the 27 features, the first 9 features are the energy potentials related to alphahelices (H), the next 9 features are related to betastrands (E) and the last 9 features are related to coils (C) as seen in Fig. 1 and 2. This gives a particular advantage in getting better classification accuracy, since this information can be used during the training phase (although this information will not be available on a blind set or a new set of proteins). Based on this prior knowledge, class specific features of the target class can be given extra weights (importance) compared to the rest of the features that belong to the negative classes. Hence the class specific features of each class (9 columns per class) were scaled (values boosted) according to a predetermined factor prior to building a training model. These factors (not unique) were obtained by brute force trial and error method, where selection was based on getting better classification results. It is noteworthy that the classification accuracy after this scaling depends on the scaling factors used, and ranges from 60% (for non-scaled data or data scaled with sub-optimal boosting values), to over 95%, when the optimal scaling factors are used. The first 9 features of all samples belonging to the H class, were scaled by a factor of 5, while the second set of 9 features were scaled by a factor of 3 and the last set of 9 features were scaled by a factor of 8. The scaling of data improves the classification accuracy considerably during the training phase. Samples which were scaled according to their classes were used for the 10-fold cross-validation in WEKA (Witten and Frank, 2005), which gave very high results for SVM and Naïve Bayes algorithms. Since it is not possible to perform class-specific feature scaling during testing (blind) phase for the ELM method, three sets of test samples were generated for each sample in the test set. The first set had the first 9 features boosted in the same ratio as for the H class for all samples. The second set of test samples had the next set of 9 features boosted according to the factor used for the E class for all samples and the third set of test samples had the last set of 9 features scaled according to the factor used for the C class

for all samples. Each test set was sent in turn and the votes were collected for the classification. For robustness, ten sets of training models were used to get the classification results for the same test set. Each training model yielded a set of three votes for each sample. These votes were all gathered to determine the class which receives the maximum number of votes. The results for the classification accuracies with and without feature scaling (value boosting) are given in the results section. Blind testing with voting was not done for SVM and Naïve Bayes algorithms since it would require modification of WEKA code.



Figure 1: Visualization of data without feature scaling. Energy potentials are represented along the x-axis, the first nine features belong to helix (H), the next 9 features are for strand (E) and the last set of features 19 - 27 for coil (C). The color intensity indicates the value of the potential energy, with a dark blue for a low value and a red indicates a high value. The residues (total: 4282) along the y-axis have been sorted according to the three classes, where residues 1 - 1487 belong to class H, 1488 - 2541 belong to class E and 2542 - 4282 belong to class C. Note: there is not much horizontal differentiation among the three classes which becomes evident in Fig. 2, after data is subjected to feature specific scaling. Results for classification of this unscaled data is given in Table 1.

#### 3.1 Two Stage Extreme Learning Machine

The ELM-PSO consists of the Extreme Learning Machine (ELM) classifier as the main algorithm, which uses a set of training samples to build a model. During the training phase, PSO is called upon to optimize the parameters, such as weights, number of hidden neurons and bias of the ELM, which results in improved classification accuracy. These parameters are stored and used during the testing phase. ELM is an improved version of a feed-forward neural network consisting of a single hidden layer. The initial set of input weights are chosen randomly, but they are tuned later by the PSO. The output weights from the hidden layer to the output layer are analytically calculated, using a pseudo inverse. A sigmoidal activation function is used for the hidden layer and a linear activation function is used for the output neurons. Huang, Zhu and Siew (2006) give a comprehensive discussion of ELM. The ELM algorithm consists of the following steps:

- 1. Select the number of hidden neurons (H) and a suitable activation function for a given problem.
- 2. Randomly choose the input weight (W) and bias (b).
- 3. Analytically calculate the output weight *using a pseudo inverse* which *speeds up* the traditional neural network algorithm tremendously.
- 4. Store the calculated weights (W, b) and hidden neurons (H) which yield the best training results.
- 5. Use these stored values for estimating the class label during testing phase.

The estimated class label  $C_i$  is calculated using equation (1) where  $y_i^k$  is the neural network output for each class k, for sample *i*.

$$\hat{c}_i = \arg \max_{k=1,2,...,C} y_i^k$$
 (1)

An improved version of the ELM algorithm proposed by Saraswathi and Suresh et al., (2010), shows that a random selection of initial parameters (W, b, H) affects the performance of the ELM classifier significantly. Tuning of input parameters using PSO, improves classifier performance considerably, by minimizing the error (Eq. 2), which is the distance between the neural network output (Y) and the target classes (T).

$$H^*, W^*, b^* \} = \arg \min_{H, V, b} \{Y - T\}$$
 (2)

#### 3.2 Particle Swarm Optimization

A stochastic optimization technique called Particle Swarm Optimization (PSO) was developed by Clerc and Kennedy (2002). This method mimics the intelligent social behavior of flocks of birds or schools of fish, represented as particles in a population. These particles work together to find a simple and optimal solution to a problem in the shortest possible time. The PSO algorithm is initialized with a set of random solutions called particles. The algorithm iteratively searches a multidimensional space for the best possible solution, determined by a fitness criterion. PSO will find the *best combination* of hidden neurons, input weights, and bias values and return the (training) validation efficiency obtained by the ELM algorithm along with the best ELM parameters to obtain *better* generalization performance. The *best parameters* are stored and used during the testing phase.

### **4 RESULTS AND DISCUSSION**

Several training models were built using ELM and two other algorithms, namely SVM and Naïve Bayes (NB) from the WEKA (Witten and E. Frank, 2005) suit of software for data classification. A 10-fold cross validation was performed for SVM and NB, where 90% of the proteins were used to build the training model while the remaining 10% were retained for testing the model, but all input information was scaled according to previously described values. A blind test was conducted using ELM with 4797 proteins for training and 4835 for testing. These residues were selected from a random selection of 30 proteins for the training set out of 400 proteins, while the test samples came from a separate set of 41 proteins retained for testing. Preliminary studies for the ELM-PSO classifier, SVM and NB show high accuracies of around 99% for the scaled training as seen in Table 2, while the results for the unscaled version of the data, as seen in Table 1, is much lower at only ~60% or less. The unscaled version of the data uses only row specific feature information while the scaled data also uses column specific class information which increases the accuracy considerably.

The lower testing accuracy of 94.4 % for the ELM (blind) tested 4835 samples might be due to the smaller number of residues tested as compared to the other two models built from SVM and NB with the full data set. The ELM classifier trains on sets of 2000 to 3000 samples at a time and builds several of these models by selecting samples at random from the pool of available training samples (from the 400 training proteins), a very computationally intensive process. The parameters for every ELM model are optimized by calling PSO and a single pattern from the test set is repeatedly tested by each model, giving a consensus classification for the type of the test sample. The class that occurs with the highest frequency in these classifications is taken to be the predicted class for this test sample. Preliminary results for a set of 4835 test samples are given in Table1 and Table 2 for scaled and unscaled data.

On the other hand the high accuracies for SVM and NB can be attributed to the technique of cross validation where the input data is uniformly scaled according to previous criteria, using feature specific class information, which results in higher accuracy. There is *no blind test of data*. So, unless the algorithm can discern this feature specific pattern automatically without involving the computationally intensive ELM-PSO method that was used here, it is not very practical. Future work will aim to improve the ELM-PSO algorithm to learn this information automatically.

Table 3 shows that the ELM-PSO methods perform very well compared to other studies in the literature for scaled data. The accuracy on the *unscaled* data is lower for all models and is comparatively low for the blind test, indicating that the learning algorithm needs further tuning to discern the column-wise information during (blind) testing phase. The column-wise class information is a unique feature of our data that separates the three classes linearly and hence gives high results. Table1 and Table 2 also give the F-measure and area under the curve (AUC) values for SVM and Naïve Bayes classifications. These terms help us to gauge the quality of the predictions.



Figure 2: Visualization of the same sample data shown in Figure 1, given here with feature scaling. Descriptions of the X,Y axes and colors are the same as given in Figure 1. Compared to Figure 1, it can be seen that class-specific feature scaling provides for a distinct separation of the classes, which results in higher accuracy during classification, using ELM, SVM-SMO and Naïve Bayes algorithms, with results shown in Table 2.

The performance of classifications can be evaluated in terms of the true positives (TP-correct) and false positive (FP-error) terms. Similar definition holds for true negatives (TN) and false negatives (FN). The output of a classification might provide estimated probabilities which determine the predicted class according to a pre-set threshold. TP rate and FP rate can be graphed as coordinate pairs which form the receiver operating characteristic curve (ROC curve).

The area under this ROC curve (AUC or AUROC) helps to aggregate the performance of all the testing results, where a higher value closer to 1.00 denotes perfect performance. F-measure gives the test's accuracy. It uses *precision* p and *recall* r of the test, where p is the ratio of correct results divided by *all returned results* (TP/(TP+FP)) and r is the number of correct results divided by the number of *expected* results (TP/(TP+FN)). F-measure is calculated as given in equation (3), where the best score for F-measure can be as high as 1 and the worst score can be as low as 0.

$$F_{measure} = \frac{2*(\text{precision} * \text{recall})}{\text{precision} + \text{recall}}$$
(3)

Table 1: Confusion matrix and accuracies for the three classes of secondary structures, for data *without feature scaling*, using ELM-PSO, SVM and Naïve Bayes.

Confusion Matrix - ELM-PSO - without feature scaling							
	Н	E	С	% correct			
Н	1147	116	457	66.7 QH			
Е	300	329	474	27.1	QE		
С	604	175	1195	30.6	QC		
			Total	4797			
				55.7	Q3		
C	onfusion I	Matrix – S	VM – wit	hout feature sca	aling		
C	onfusion M	Matrix – S E	VM - wit	hout feature sca % correct	aling		
Co H F	H 3153 817	Matrix – S E 533	VM – wit C 1672	hout feature sca % correct 58.8 22.8	aling QH OF		
Co H E C	H 3153 817 1446	Matrix – S E 533 1353 595	VM – wit C 1672 1411 5083	hout feature sc. % correct 58.8 22.8 20.5	aling QH QE QC		
C H E C	H 3153 817 1446	Matrix – S E 533 1353 595	VM – wit C 1672 1411 5083 Total	hout feature sc. % correct 58.8 22.8 20.5 16063	aling QH QE QC		
Co H E C	H 3153 817 1446	Matrix – S E 533 1353 595	VM – wit C 1672 1411 5083 Total	hout feature sc. % correct 58.8 22.8 20.5 16063 59.7	Aling QH QE QC Q3		
C H E C	H 3153 817 1446	Matrix – S E 533 1353 595 F-Me	VM – wit C 1672 1411 5083 Total easure	hout feature sc. % correct 58.8 22.8 20.5 16063 59.7 58.5	QH QE QC Q3		

	Н	Е	С	% correct	
Н	3244	1217	897	60	QH
Е	705	2168	708	60	QE
С	2028	2168	708	47.6	QC
			Total	16063	
				54.8	Q3
		F-Measure		55.1	
		AUC		73.5	

Table 2:	Confusion	matrix	for	the	three	classes	of
secondary	structures,	for data	with	feat	ure sca	aling, u	sing
ELM-PSO	, SVM and	Naïve Ba	aves.				

Confusion Matrix - ELM-PSO - with feature scaling								
	Н	Е	С	% correct				
Н	1814	0	0	100	QH			
Е	56	942		94.3	QE			
С	224	0	1799	89.9	QC			
			Total	4835				
				94.4	Q3			
	Confusion Matrix – SVM - with feature scaling							
	Н	Е	С	% correct				
Н	24854	67	8	99.7	QH			
Е	0	16879	4	100	QE			
С	0	0	31096	100	QC			
		Total		72908				
_			í a	99.9	Q3			
	F- Measure 99.8							
	AUC 99.9							
Confusion Matrix – Naïve Bayes - with feature scaling								
ושם		E	= C	% correct	Z			
Η	24896	33	0	99.9	QH			
E	256	16627	0	98.5	QE			
С	0	19	31077	99.9	QC			
			Total	72908				

Table 3: Comparison of results for secondary structure prediction using ELM-PSO - feature scaled data, with other studies in literature.

F- Measure

AUC

99.6

99.6 100 Q3

Method	Q3 (%)	QH (%)	QE (%)	QC (%)
PHD (Rost and Sander, 1999)	70.8	72.2	66.0	72.0
JNet server (Cuff and Barton, 2000)	76.4	78.4	63.9	80.6
SVMpsi (Kim and Park, 2003)	76.6	78.1	65.6	81.1
SPINE server (Dor and Zhou, 2007)	80.0	84.44	72.23	80.46
ELM-PSO with feature scaling	94.4	100	94.3	89.9

## **5** CONCLUSIONS

A two stage approach for secondary structure prediction was presented where an Extreme Learning Machine (neural network) was used along with Particle Swarm Optimization (ELM-PSO) for classifying a reduced set of three secondary structures, namely, alpha-helix, beta-strand and coil. The data was generated using CABS potential energy. ELM-PSO needs improvement to achieve better accuracies on blind tests so that comparative results can be achieved on new proteins.

### ACKNOWLEDGEMENTS

We thank Pawel Gniewk, a student at 'Theory of Biopolymers, Faculty of Chemistry, Warsaw University', whose original idea and algorithm was used to generate the potentials data that was used for the secondary structure predictions. We acknowledge the support of National Institutes of Health through grants R01GM081680, R01GM072014, and R01GM073095 and the support of the NSF grant through IGERT-0504304.

### REFERENCES

Altschul, S., Madden, T., and Schaffer, A., 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucl Acids Res*, 25, 3389 – 3402.

4ND

- Clerc, M. K. and Kennedy, J., 2002. The particle swarm explosion, stability, and convergence in a multidimensional complex space. *IEEE Trans olutionary Comput*, 6 (1) 58-73.
- Chou, P. Y. and Fasman, G. D., 1974. Prediction of protein conformation. *Biochemistry*, 13(2), 222-245.
- Cole, C., Barber, J. D., and Barton, G. J., 2008. The Jpred 3 secondary structure prediction server. *Nucleic Acids Research*, 36 (Web Server issue): W197–W201.
- Cuff, J. A. and Barton, G. J., 2000. Application of multiple sequence alignment profiles to improve protein secondary structure prediction. *Proteins*, 40(3), 502-511.
- Dor, O. and Zhou, Y., 2007. Achieving 80% Ten-fold Cross-validated Accuracy for Secondary Structure Prediction by Large-scale Training. *PROTEINS: Structure, Function, and Bioinformatics*, 66, 838-845.
- Garnier, J., Osguthorpe, D. J. and Robson, B., 1978. Analysis of the accuracy and implications of simple methods for predicting the secondary structure of globular proteins. *J Mol Biol*, 1, 97-120.
- Garnier, J., Gibrat, J. F., and Robson, B., 1996. GOR secondary structure prediction method version IV. *Methods Enzymol*, 226, 540-553.
- Huang, G. B., Zhu, Q. Y., and Siew, C. K., 2006. Extreme learning machine: Theory and applications. *Neurocomputing*, 70(1-3), 489-501.
- Jones, D., 1999. Protein secondary structure prediction based on position-specific scoring matrices. J Mol Biol, 292, 195 – 202.

- Kabsch, W. and Sander, C., 1983. Dictionary of protein secondary structure: pattern recognition of hydrogenbonded and geometrical features. *Biopolymers*, 22(12), 2577-2637.
- Kihara, D., 2005. The effect of long-range interactions on the secondary structure formation of proteins. *Prot Sci.*, 14(8), 1955–1963.
- Kim, H. and Park, H., 2003. Protein Secondary Structure Prediction Based on an Improved Support Vector Machines Approach. *Protein Eng*, 16, 553-560.
- Kloczkowski, A., Ting, K. L., Jernigan, R. L., and Garnier, J., 2002. Combining the GOR V algorithm with evolutionary information for protein secondary structure prediction from amino acid sequence. *Proteins*, 49, 154-166.
- Kolinski A., 2004. Protein modeling and structure prediction with a reduced representation. *Acta Biochim Pol*, 51, 349-371.
- Lomize, A. L., Pogozheva, I. D. and Mosberg, H. I., 1999. Prediction of protein structure : The problem of fold multiplicity. *Proteins*, 37, 199-203.
- Montgomerie, S., Sundaraj, S., Gallin W., and Wishart, D., 2006. Improving the Accuracy of Protein Secondary Structure Prediction Using Structural Alignment. *BMC Bioinformatics*, 7, 301.
- Ortiz, A. R., Kolinski, A., Rotkiewicz, P., Ilkowski, B. and Skolnick, J., 1999. Ab initio folding of proteins using restraints derived from evolutionary information. *Proteins Suppl 3 (CASP3 Proceedings)*, 177-185.
- Pollastri, G., Martin, A., Mooney, C. and Vullo, A., 2007. Accurate prediction of protein secondary structure and solvent accessibility by consensus combiners of sequence and structure information. BMC Bioinformatics, 8(1), 201.
- Qian, N. and Sejnowski, T. J., 1988. Predicting the secondary structure of globular proteins using neural network models. *J Mol Biol*, 202, 865-884.
- Rost, B. and Sander, C., 1993. Prediction of protein secondary structure at better than 70% accuracy. J. Mol. Biol., 232, 584–599.
- Rost, B., 2001. Review: Protein Secondary Structure Prediction Continues to Rise. J Struct Bio, 134, (2-3), 204-218.
- Rost, B., Yachdav, G. and Liu, J., 2004. The PredictProtein Server, *Nucl Acids Res*, 32, Web Server issue, W321-W326.
- Saraswathi, S., Suresh, S., Sundararajan, N., Zimmerman, M. and Nilsen-Hamilton, M., 2010. ICGA-PSO-ELM approach for Accurate Cancer Classification Resulting in Reduced Gene Sets Involved in Cellular Interface with the Microenvironment. *IEEE Transactions in Bioinformatics and Computational Biology*, http://www.computer.org/portal/web/csdl/doi/10.1109/ TCBB.2010.13.
- Ward, J. J., McGuffin, L. J., Buxton, B. F. and Jones, D. T., 2003. Secondary structure prediction with support vector machines. *Bioinformatics*, 19(13), 1650-1655.
- Witten, I. H. and Frank, E., 2005. Data Mining: Practical machine learning tools and techniques, (2nd ed.) San Francisco: Morgan Kaufmann.