Amino Acid and Proximate Compositions of Cultured and Wild Oreochromis niloticus (Linnaeus 1758) from Makurdi-Nigeria

A.S. Nege¹ and E.T. Akange²

¹Graduate Student; Fisheries and Marine Biotechnology, Universitas Airlangga, 60286, Surabaya Indonesia ²Fisheries and Aquaculture Department, University of Agriculture, P.M.B 2373, Makurdi, Nigeria

Keywords: Amino acids, Cultured, Makurdi, Oreochromis niloticus, Proximate compositions, Wild.

Abstract: This study was aimed at identifying the fish habitat that harbors *O. niloticus* with higher nutrients and availing these findings for nutritional guidance. Amino acid and proximate compositions of *O. niloticus* from Lower Benue River and UAM fish farm were determined monthly for a 3month period, using methods of Benitez (1989) and AOAC (2006) respectively. The mean of Glutamic acid (12.51±0.64 and 11.85±0.67) and Alanine (5.40±0.23 and 4.50±0.41) varied significantly (p<0.05) between the wild and cultured while other amino acids were not. In October, ash (4.62±0.02 and 3.69±0.02), fat (4.71±0.06 and 3.61±0.02), protein (14.62±0.07 and 9.98±0.02) and moisture (75.67±0.07 and 79.65±0.15) varied significantly between the wild and cultured *O. niloticus*. Crude protein (20.46±0.01 and 18.75±0.04) and moisture (69.32±0.02 and 73.35±0.20) significantly varied in November between the samples respectively. Only crude protein (18.89±0.04 and 20.31±0.06) varied statistically in December between wild and cultured. The mean of crude protein (17.99±1.22 and 16.34±1.86) and the mean moisture (72.53±1.16 and 74.94±1.52) differed significantly between the wild and cultured. From these results. *O. niloticus* wild expressed superiority over the cultured, however, both River Benue at Makurdi and the UAM fish farm harbor nutritious *O. niloticus* which is good for human consumption based on our daily amino acid and protein needs.

1 INTRODUCTION

More recently, fish has become a favorite foodstuff for many people due to several health reasons (Ali and Kiumars, 2010). However, considering the maximum utilization and the knowledge of fish composition, conducting studies on fish flesh vital in the fishery industry.

According to Silva and Chamul (2000), the nutritional composition of fish varies greatly from one species and individual to another depending on numerous factors such as age, feed intake, sex, the environment and season. Puwastien et al., (1999) stated that fish proximate composition serves as a good indicator of fish quality, and it varies with parameters like genetic strain, diet, feed rate and age.

In order to flourish and maintain bodily functions, fish similarly to other animals require different nutrients in sufficient quantities (Ashraf et al., 2011). It also has the ability to synthesize some but not all nutrients. Hence, some of these nutrients must be obtained from outside sources. Apart from the natural productivity of ponds, cultured fish is provided with nutrient-rich feed, while captured fish on the other hand, solely depends on natural food in the aquatic environment for its survival and sustenance. These variations have been reported to directly related to the growth, health and body composition of fish. Therefore, fish composition is a good index of fish food availability and feeding habits (Ashraf et al., 2011).

For the nature of the raw material in fish to be known before chilling, freezing, smoking orcanning can be correctly applied. Fish processors are directly interested in the proximate composition of fish (FAO, 2004).

Bakir et al., (1993) mentioned that *Oreochromis* niloticus, Tilapia zilli, Sarotherodon galilaeus, as well as *Clarias gariepinus*, *Clarias angullaris* and *Heterobranchus longifilis* from the Cichlidae and Clariidae families form part of the freshwater fish species mostly cultured in the developing nations. Hence, the need for using cultured and wild O. niloticus for this study.

Nege, A. and Akange, E.

Amino Acid and Proximate Compositions of Cultured and Wild Oreochromis niloticus (Linnaeus 1758) from Makurdi-Nigeria. DOI: 10.5220/0007547105670570

In Proceedings of the 2nd International Conference Postgraduate School (ICPS 2018), pages 567-570 ISBN: 978-989-758-348-3

2 MATERIALS AND METHODS

2.1 Sample Collection

Twelve (12) samples of O. niloticus were collected for this study, comprising of six (6) samples each from Lower Benue River at Wadata/Tse-Adee landing site Makurdi (with average weight of 70.13g) and the University of Agriculture Makurdi (U.A.M) fish farm (with average weight of 67.42g). The samples were then taken to the Hydrobiology and Fisheries Research laboratory, University of Jos, Plateau State, Nigeria where the laboratory analyses were conducted.

2.2 Study Duration

The duration for this study was three (3) months, and the laboratory analysis of fish carcass was carried out in a monthly basis from October to December, 2014.

2.3 Amino Acid Profile Determination

The Amino acid profile in the known sample was determined using methods described by Benitez (1989) in Technicon sequential Multi-Sample Amino Acid Analyzer (TSM).

2.4 Determination of Proximate Composition

Proximate composition of moisture, crude protein, fat, and ash was determined based on the method described by the Association of Official Analytical Chemists (AOAC, 2006). All values of proximate parameters were converted and presented on wet weight basis after each calculation.

2.5 Statistical analysis

Student t-test and descriptive statistics (mean and standard error of the means) were used to analyze the data obtained from this work.

3 RESULTS

A comparison between the amino acid profiles of *O. niloticus* from Lower Benue River and that of U.A.M fish farm indicated a significant difference (p<0.05) between Glutamic acid (12.51±0.64 from the river and 11.85±0.67 from the pond) and also, Alanine $(5.40\pm0.23$ from the wild and 4.50 ± 0.41 from the cultured) while the rest amino acids remained insignificantly different (p<0.05) as shown in Table 1.

Table 1: Amino acid profiles of O.	niloticus from Lower
River Benue and U.A.M fish farm	

	Source					
Amino Acid (g/100g protein)	Lower River Benue	UAM Farm	P-Value			
Lysine	6.84±0.28	6.78±0.28	0.681			
Histidine	2.07±0.13	2.03 ± 0.08	0.267			
Arginine	5.97±0.21	5.29±0.21	0.681			
Aspartic acid	8.96±0.19	8.47±0.31	0.732			
Threonine	2.52±0.22	2.31±0.15	0.851			
Serine	4.08±0.15	3.52±0.25	0.623			
Glutamic acid	12.51±0.64	11.85±0.67	0.047*			
Proline	4.21±0.24	3.81±0.34	0.241			
Glycine	6.59±0.22	5.47±0.37	0.369			
Alanine	5.40±0.23	4.50±0.41	0.047*			
Cystine	0.79 ± 0.07	0.73 ± 0.06	0.091			
Valine	4.12±0.29	3.53±0.27	0.235			
Methionine	2.26±0.09	2.19 ± 0.06	0.414			
Isoleucine	3.44±0.16	3.15±0.09	0.364			
Leucine	6.87±0.15	6.51±0.22	0.259			
Tyrosine	2.89±0.15	3.00 ± 0.09	0.871			
Phenylalanine	3.81±0.13	3.65±0.21	0.681			
*indicates statistical difference						

(p<0.05). Source: Laboratory work.

Table 2 shows the results of monthly proximate compositions of *O. niloticus* from Lower Benue River and that of U.A.M fish farm. It reveals that all proximate parameters varied significantly (p<0.05) during the month of October with fat (4.71 ± 0.06 and 3.61 ± 0.02), ash (4.62 ± 0.02 and 3.69 ± 0.02), crude protein (14.62 ± 0.07 and 9.98 ± 0.02) and moisture (75.67 ± 0.07 and 79.65 ± 0.15) for *O. niloticus* from Lower Benue River and that of U.A.M fish farm respectively.

During the month of November, fat and ash did not vary significantly while crude protein and moisture varied significantly at p<0.05 with the following values; 20.46 \pm 0.01 and 18.75 \pm 0.04 crude protein and moisture content of 69.32 \pm 0.02 and 73.35 \pm 0.25 of *O. niloticus* from the river and fish farm respectively as shown in Table 2. In December, the results showed that only crude protein (18.89 \pm 0.04 and 20.31 \pm 0.06) varied significantly between wild and cultured *O. niloticus* while the other parameters were not statistically different (p<0.05) as equally showed in Table 2.

Variations in all the proximate parameters for the entire study period (Oct. to Dec.) shows that the mean of crude protein $(17.99\pm1.22 \text{ and } 16.34\pm1.86)$ and the mean of moisture $(72.53\pm1.16 \text{ and } 16.34\pm1.86)$

74.94 \pm 1.52) differed significantly (p<0.05) while fat and ash were not significantly different through the

period between *O. niloticus* from Lower Benue River and U.A.M fish farm as shown in Figure 1.

Month		Fraction			
	Source	Fat	Ash	Protein	Moisture
October	Lower River Benue	4.71±0.06	4.62±0.02	14.62±0.07	75.67±0.07
	UAM Farm	3.61±0.02	3.69±0.02	9.98±0.02	79.65±0.15
	P-value	0.039*	0.035*	0.013*	0.039*
November	Lower River Benue	2.88±0.02	1.62±0.02	20.46±0.01	69.32±0.02
	UAM Farm	3.12±0.07	2.07±0.06	18.75±0.04	73.35±0.25
	P-value	0.056	0.058	0.037*	0.030*
December	Lower River Benue	2.89±0.08	4.65±0.04	18.89±0.04	72.59±0.25
	UAM Farm	3.39±0.04	4.16±0.04	20.31±0.06	71.83±0.01
	P-value	0.068	0.079	0.041*	0.160

Source: Laboratory work.

Table 2: Monthly Variation of Proximate compositions of O. niloticus from Lower Benue River and U.A.M fish farm

*indicates statistical difference (p<0.05).

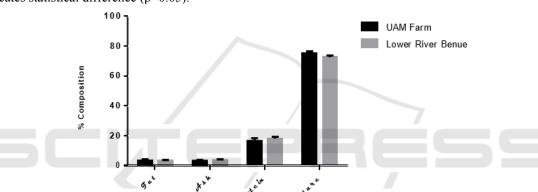


Figure 1: Mean proximate compositions of O. niloticus from Lower Benue River and U.A.M fish farm

Fractions

4 **DISCUSSION**

The mean ash values for *O. niloticus* from Lower Benue River (3.63 ± 0.59) and U.A.M (3.31 ± 0.48)

did not fall in same range with the results obtained by Osibona et al., (2009) from *Tilapia zilli* $(1.2\pm0.2\%$ ash). This difference may be due to environmental factors and variation in the ages of fish samples used. Crude protein result of *O. niloticus* from Lower Benue River (17.99±1.22) was higher than the crude protein value of *O. niloticus* from the wild (14.328 Cp) studied by Ayeloja et al., (2013) but lower than that of wild Tilapia zilli (19.0±1.9) obtained by Osibona et al., (2009). When the crude protein of *O. niloticus* from Lower Benue River (17.99±1.22) and U.A.M (16.34±1.86) are compared, there was a variation of 1.65%. However, all the values of mean crude protein from this study lie within the range obtained from the findings of Eyo (2001) which was between 15-20% crude proteins in fresh fish tissues. This indicates that fish, irrespective of their species and source of capture, have a certain range for their crude protein content.

When the mean moisture value of *O. niloticus* from Lower Benue River is compared with that of U.A.M, it indicated a significant variation (p<0.05) which clearly justifies the fact that fish from same species obtained from different environments could vary in term of their nutrient contents. The mean of moisture contents of *O. niloticus* from Lower Benue River and U.A.M were all notably less than the moisture content of *O. niloticus* obtained by Eyo. This could be due to difference in geographical location. *Oreochromis niloticus* obtained from Lower Benue River had moisture content of 72.53±1.16. When compared with the 78.325%

moisture obtained from wild *O. niloticus* by Ayeloja et al., (2013), there was a difference of 5.80%. This difference could probably be due to differences in fish age, feed intake, location, and even sexes as reported by Silva and Chamul (2000) and Eyo (2001).

In terms of fat, the results of Ramlah et al. (2016) from Indonesian wild and cultured *O. niloticus* (0.10 and 0.18 respectively) were far lower than fat contents of this study $(3.49\pm0.387 \text{ and } 3.37\pm0.092 \text{ respectively})$.

The crude protein of wild *O. niloticus* (18.46 ± 1.22) from this study was higher compared to that of wild

O. niloticus from Indonesia which was 12.94 as reported by Ramlah et al (2016). However, the cultured O. niloticus from this study which had 15.87 ± 1.86 is 0.92% less than the 16.79 obtained from cultured Indonesian *O. niloticus* by Ramlah et al. (2016).

The variations in amino acid content and proximate compositions in this study were possibly due to differences in capture environments, months of the year, the food availability, sexes, age and sizes of the sample fishes used.

5 CONCLUSIONS

This research clearly reveals the superiority in nutritional composition of wild *O. niloticus* obtained from Lower Benue River at Makurdi against its counterpart from the ponds of University of Agriculture Makurdi fish farm. However, both the wild and cultured *O. niloticus* from Makurdi are good for human consumption since they are all high in protein and amino acid contents, and their nutritional values fall within ranges established by other authors.

REFERENCES

- Ali, A. and Kiumars, P. (2010); Chemical and proximate composition properties of different fish species obtained from Iran. World Journal of Fish and Marine Science, 2:237 239.ISSN 20784589.
- AOAC (Association of OfficialAnalytical Chemists), (2006); Official Method of Analysis of the AOAC (W. Horwitz Editor, Eighteenth Edition, Washington; D. C., AOAC). In AOAC International, Suite 500, 481North Frederick Avenue, Gaithersburg, Maryland 20877 2417, USA.
- Ashraf, M., Abdul Rauf, Asma Zafar and Shahid Mehboob (2011); Nutritional value of wild and cultivated Silver

(Hypophthalmichthys molitrix) and Grass carp (Ctenopharyngodo idella). International Journal of Agriculture and Biology, Vol.13, PP.210 214.

- Ayeloja, A. A.; George, F. O. A.; Dauda, T. O.; Jimoh, W. A. and Popoola M. A (2013); Nutritional comparison of captured Clarias gariepinus and Oreochromis niloticus. International Research Journal of Natural Sciences. Vol. 1, No. 1, pp. 9-13.
- Ayyappan, S; Jena, JK; Gopalakrishnan, A. and Pandey, Ak (2006); Handbook of Fisheries and Aquaculture. Indian Council of Agricultural Research New Delhi. ISBN:81-7164-061-3.
- Bakir, H. M., Melton, S.L. and Wilson, J.L. (1993); Fatty acid composition, lipids and sensory characteristics of white amur (Ctenopharyngodon idella) fed different diets. J. Food Sci. 58(1):90-95.
- Benitez, L. V. (1989); Amino Acid and fatty acid profiles in aquaculture nutrition studies, p. 23- 35.In S.S. De Silva (ed.) Fish Nutrition Research in Asia. Proceedings of the Third Asian Fish Nutrition Network Meeting. Asian fish Society Special Publication.4, 166 p. Asian Fisheries Society, Manila, Philippines.
- Eyo, A. A (2001); Fish Processing Technology in the Tropics. Published by National Institute for Freshwater Fisheries Research (NIFFR), P.M.B 6006, New Bussa, Nigeria.
- Food and Agricultural Organization, (2004); The composition of fish. Accessed from http://www.fao.org/wairdoes/tx5916e/×5 16co1.htm., pp: 1-80.
- Osibona, A. O.; Kusemiju, K. and AkandeG. R (2009); Fatty acid composition and amino acid profile of two freshwater species, African catfish (Clarias gariepinus) and Tilapia (Tilapia zillii). African Journal of Agriculture Food Nutrition and Development, Vol. 9 No. 1 pp. 608 621.
- Puwastien, P.; Raroengwichit, M.; Sungpuag, P. and Judprasong, K. (1999); "Thai Food Composition Tables, 1st Edition". Institute of Nutrition, Mahidol University (INMU), Thailand Asean foods Regional Database Centre of INFOODS.
- Ramlah, Eddy Soekendarsi, Zohrah Hasyim and Munis Said Hasan (2016); Comparison of Nutritional content of Tilapia Oreochromis niloticus from Mawang's Lake Gowa and Hasanuddin University Lake Makassar city. In: Jurnal Biologi Makassar (BIOMA), Vol.1, No.1.
- Silva, J.J. and Chamul, R.. S (2000); Composition of marine and fresh water finfish and shellfish species and their products. In: Martin, R.E., E.P. Carter, E.J. Flick and L.M. Davis (Eds.), Marine and fresh water products handbook, Lancaster, Pennsylvania, U.S.A: Technomic Publishing Company, pp: 31-46