A Computer Vision Approach to Fertilizer Detection and Classification

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Abstract: This paper introduces a computer vision based pipline for the classification of different types of fertilizers from collected images. For robust boundary detection of individual corns in a heap of grains, we used YOLO11 for classification and Segment Anything 2 for segmentation in an active learning fashion. The segmenter as well as the classifier are iteratively improved starting with an initial set of handcrafted training samples. Despite the high diversity in grain structures, the relatively simple camera setup and the limited number of handcrafted training samples, a classification accuracy of 99.996% was achieved.

1 INTRODUCTION

Many application areas greatly benefit from the use of cutting-edge Artificial Intelligence (AI) technologies. Applying intelligent systems often result in precise manufacturing, improved product quality, faster production as well as reduced costs. This success is now drawing increasing attention to its potential in agriculture, where AI can play a pivotal role in modernizing traditional farming practices. In particular the use of Computer Vision (CV) combined with stateof-the-art Machine Learning (ML) supports the digital transformation of age old-farming to smart agriculture (Ghazal et al., 2024), (Dhanya et al., 2022). Applications of AI range from weed and crop classification (Adhinata et al., 2024), leaf disease detection (Sarkar et al., 2023), to the weight prediction of livestock such as pigs (Xie et al., 2024).

The pipeline presented in this paper is part of an intelligent system designed to identify key characteristics of fertilizers. According to the EU Fertilizer Products Regulation (FPR), farmers must adhere to strict regulations regarding environmental and safety requirements. In particular, they are required to document the type and amount of fertilizer applied, as well as the concentrations of total nitrogen and phosphate. Farmers must also ensure minimum distance from waters and public pathways. With the worldwide use of fertilizers increasing to 195.38 million tons in 2021 (Figure 1), 90% of fertilization costs are attributed to the fertilizer itself. In the past, farm-



Figure 1: Global consumption of agricultural fertilizer from 1961 to 2022, by nutrient (in million metric tons). Data from: International Fertilizer Industry Association (ifas-tat.org).

ers had limited fertilizer options, but today they must deal with thousands of products (Figure 2), including blends and biofertilizers. Fertilizer types can be distinguished by physical characteristics such as density, volume, and granularity.

Rather than relying on a variety of sensors to extract these characteristics, we propose a simple and straightforward hardware setup for image aquisition, followed by a state-of-the-art CV-ML pipeline. This pipeline is built using YOLO11 (Redmon et al., 2015) and Segment Anything 2 (Ravi et al., 2024). Models are trained using only a small set of handcrafted sam-

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ples in an active learning-based approach.



Figure 2: Cuttings from images of the 16 fertilizers used in the project.

2 SYSTEM DESIGN

The motivation for our system design is an industrial requirement to develop a cost-effective method for testing and detecting fertilizers with as little manpower needed as possible. This also includes the development of a testing device for the controlled and standardized recording of photos. The presented subsystem is part of a bigger research project aiming to create an application that enables fertilizer samples to be analyzed and, if necessary, measured under laboratory conditions.

Hardware Setup 2.1



(a) Front view

Figure 3: Imaging box.

A controlled environment is essential for obtaining reproducible results, particularly in CV applications where consistent image capture is key. To achieve this, we designed a small, portable, and cost-effective imaging box. The box is constructed with an aluminum frame, white cardboard side panels, and a wooden lid, which together provide a nearly lighttight seal to minimize external light interference (Figures 3(a),3(b)). The bottom is sealed with a wooden plate that includes a cutout for a sample cup (Figure 4). With a radius of 87.5 mm, the sample cup is 3D-



Figure 4: Sample cup.

printed from PLA and divided into three sections: two crescent-shaped sections and one nearly rectangular section. The division can be adjusted depending on the specific needs. This setup allows for capturing images of multiple specimens simultaneously, speeding up the image acquisition process. In our specific application it enables to take images of multiple fillings of the same sample at once. The plate also contains a mounting bracket to securely hold the sample cup in place, while allowing it to be rotated if needed. To illuminate the interior of the box, we installed four independently controllable LED light bars, each providing 485 lumens with a color temperature of 4000K (Figure 6). The light bars emit diffuse light, which, in combination with the white cardboard side panels, ensure even illumination throughout the imaging box. Additionally, the angle of the lights can be adjusted as needed to achieve optimal lighting conditions. For image capture, we used a Raspberry Pi HO Camera Module equipped with the default 6mm lens, connected to a Raspberry Pi 4. The camera is securely mounted on an aluminum extrusion, which is fixed to the frame of the box to ensure stability and consistent positioning (Figure 5).



Figure 5: Camera closeup.

A checkerboard pattern was printed on the inside

of the bottom plate to evaluate the effect of the undistortion algorithm that will be applied (cf. Figure 9(b)). The cutout in the bottom plate (with a radius of 91.5 mm) allows the camera to capture images of the sample cup and its content, which is mounted below the box.



Figure 6: Light bar.

2.2 Software Solution for Imaging

Creating a fast, easy-to-use and expandable software solution for capturing images was a key goal of this project. For this purpose, we developed a modular, all-in-one solution that can be easily expanded and customized to meet the specific needs of any project. The software is written in Python and consists of two main components: an image acquisition module running on the Raspberry Pi, and a control module that manages the Raspberry Pi's actions. This control module runs on a local computer, which we refer to as the control computer. Additionally, the control computer receives the captured images wirelessly from the Raspberry Pi via MQTT (using Mosquitto (Light, 2017)) for further image processing. We chose MOTT because it is a lightweight, open-source protocol that is easy to set up and use (MQTT.org, 2024). It allows us to send the images to multiple destinations (e.g., the control computer and a backup server) simultaneously via an MQTT broker running on the Raspberry Pi. Our implementation includes the following key features: adjusting the camera focus (as the lens does not have autofocus), capturing images and wirelessly transmitting them to the control computer, performing white balance adjustments, and undistorting images to correct for lens distortion. Configuring the software for both the imaging box and the control computer is straightforwardly done using a simple configuation file on each device. The configuration file contains all the necessary parameters for the software to run, including the IP address of the MQTT broker, the camera's exposure time(s) and gain(s), as well as the names of the specimens to

be captured. One of the key challenges we encountered was finding a reliable solution for adjusting the camera's focus. Initially, using a live feed from the Raspberry Pi on the control computer to fine-tune the focus through manual inspection proved ineffective, as the images often appeared blurry due to imprecise adjustments. To overcome this, we developed a software module that uses a Siemens star (similar to ISO 12233:2017) and basic CV techniques to quantify the focus adjustment. A Siemens star is a test pattern used for calibrating the sharpness of a camera lens (Figure 7). Adjusting the camera's focus affects the



size of the gray area in the center of the star - the smaller the area, the sharper the image. In our setup, the star is placed on the bottom of the imaging box at the same distance from the camera as the sample cup, ensuring an accurate calibration of the focus. The process starts by capturing a single image of the star. Then a simple template matching algorithm based on 2D correlation coefficiants identifies the center of the star which is used for further calculations. Next, the camera captures a continuous stream of images and a Canny Edge Detection algorithm (Canny, 1986) is applied to detect the edges of the star. In the gray area, edge detection fails to find edges, allowing us to calculate the radius of this area using the center of the star. This radius is displayed on the screen to support the user when setting the focus to minimize the gray area for optimal sharpness (Figure 8). Once the focus is optimized, the corresponding value is saved with a timestamp for documentation. To ensure consistent colors in the images, we disabled the camera's automatic white balance and implemented a custom white balance correction method. First, we captured an image of a surface that we defined as white (e.g. a piece of paper). The average color of this image is then compared to true white (RGB: 255, 255, 255), and the resulting factor is used to correct the white balance for all subsequent images. In a final step, we used checkerboard patterns of a fixed size to apply



(a) Bad sharpness calibra- (b) Good sharpness calibration tion

Figure 8: Screenshots taken from the GUI for the sharpness calibration process.

an undistortion algorithm (Zhang, 2000), (Heikkila and Silven, 1997) for the correction of lens distortion. To ensure optimal corrections, three images of the checkerboard patterns at different orientations were used for undistortion. Figure 9 (a) and (b) show the raw image and the undistorted, centered, and whitebalance-corrected image, respectively.



Figure 9: Data preprocessing.

Afterwards to this pre-processing, we applied a simple detector for the circular markings on the edge of the sample cup. This gives us a circular cropped image. The two circular segments with the fertilizer samples are then cut out and rotated into a uniform position as shown in Figure 11. Those cropped images are later used for training and testing the ML-Pipeline.

3 CV-ML PIPELINE

For a robust detection and classification of fertilizer outlines and types, we propose a processing pipeline utilizing state-of-the-art tools, trained in an active learning approach. First, we trained a classifier to distinguish grains from other objects in the image. To accomplish this, a few grains of each fertilizer are labeled by hand in the images captured with the imaging box. These markers are defined to be rectangular bounding boxes around individual grains. With this small number of samples a first weak detector model is trained which in turn is used to process more images resulting in a bunch of new bounding boxes. Because of the weakness of the detector the bounding boxes are inaccurate (cf. red rectangle in Figure 10). To improve the accuracy, the detected bounding boxes are fed into a segmentation system with zeroshot generalization to unfamiliar objects and images. This results in a contour that encloses the grain exactly within any weak bounding box. Then the extent of the contour is used to shrink the weak bounding box to an accurate bounding box (cf. green rectangle in Figure 10).



Figure 10: Illustration of the box refining process. Red rectangle: shows the best guess of the first weak detector. Orange outline: shows the result of the zero-shot segmentation system. Green rectangle: new box defined by the limits of the outline.

In a next step, these new bounding boxes together with the available category (i.e. type of fertilizer) are used to train an improved grain detector and a classifier for the type of fertilizer. Afterwards, we applied this model to detect grains in the provided unlabeled images. Although these steps could be repeated many times to improve the accuracy, in our application one step already yields a refined model detecting very accurate bounding boxes with high classification accuracy from a small number of handlabeled samples. However, to calculate shape specific parameters of each fertilizer grain, we run the zero-shot segmentation system once again to extract outlines. To eliminate overlapping detections, a few post-processing steps are carried out.

In summary, the processing steps of our pipeline are as follows:

- 1. train a weak detector and classifier for fertilizer grains using a few hand-labled images
- 2. extract outlines and refine bounding boxes
- 3. train a refined detector and classifier for the type of fertilizer
- use refined model to detect and classify fertilizer grains of unlabeld images
- 5. extract outlines and post-process detections

In the following sections, we describe these steps in more detail.

3.1 YOLO and SegmentAnything

YOLO11 is the newest version of the well known object detection model. It is capable of real-time detection and can be easily adapted to different tasks. YOLO11 provides pretrained models for different tasks (e.g. detection, segmentation, pose estimation, etc.) which can be easily finetuned with custom data. In this paper, only the model for object detection from YOLO11 is used, which provides bounding boxes, a class label as well as a confidence score for each detected object of an image.

Because YOLO11 resamples the input images dependent on their resolution, we use SAHI (Slicing Aided Hyper Inference) (Akyon et al., 2022) for slicing the original image into overlapping slices of constant resolution. Afterwards, multiple detections whithin each image are combined using a non-maximumsuppression algorithm.

For the segmentation we use the Segment Anything Model 2 (SAM2). SAM2 is a segmentation system with zero-shot generalization that provides robust results with some known context. This context is either one or more known points inside (or outside) of the object to be segmented or else a bounding box approximating the outer bounds of the object. The latter is used in the context of this paper. As (Alif and Hussain, 2024) shown in their comprehensive review, YOLO is very well suited for applications in an agricultural context. We therefore decided in favor of the use of YOLO as detector and subsequent SAM2 for obtaining the contours, as these two tools meet the industrial requirements described at the beginning: They are easy to use and powerful enough to perform grain detection and segmentation without the need for extensive manual labeling.

3.2 Training

We conducted a total of 3 photo sessions for taking images of fertilizers as sample dataset.

Session 1. A total of 44 photos of 22 classes were taken, resulting in 88 images of circle segments. Thus, each image shows either the lower or the upper part of the filled sample cup (Figures 11(a) and 11(b)). The 22 classes consist of 16 different types of fertilizers (cf. Figure 2), 5 different types of spherical airsoft bullets (exactly 5.95 mm of diameter) and one type of plastic granulate. The six non-fertilizer classes, along

with their known parameters (e.g. diameter, roundness) have been added to improve monitoring our results. We split the dataset into two halves: 44 images, i.e. exactly two of each class, showing only the upper part of the sample cup are used in all training processes. The remaining 44 images of the lower part of the sample cup are used only once for testing.



Figure 11: Relevant regions of the filled sample cup, cropped and rotated to a uniformly position.

Session 2. A total of 46 photos of 23 classes were taken, resulting in 92 images of circle segments. In addition to session 1, we have added an additional class: small PVC tube segments in various colors with a length of 5 mm, an outer diameter of 5 mm and an inner diameter of 2.5 mm. We also replaced the camera and lens. They are the same models, but in contrast to session 1, the camera in session 2 does not have an infrared cut filter. This resulted in images with a different color spectrum. As in session 1, we again divided the data set in half and used images of the upper part of the sample cup for training and the lower part exclusively for testing.

Session 3. The same setup as in session 2, but this time we readjusted the focus, aperture and camera position and took all images of non-fertilizer classes. We then readjusted the focus, aperture and camera position and took all fertilizer images. This procedure enabled to achieve slight variations in the setup and simulate a more realistic real-world application. The images taken in this way are used exclusively for testing.

To train our first simple and thus weak grain detector we manually labeled 604 grains with fitted bounding boxes around each grain (approximately 20-30 boxes per class, 10-15 per image) using only the images taken in session 1. The original image size is 907x2435 pixels with smaller grains being only about 30 pixels wide. In general, detecting small structures in large images is challenging for many detectors. To improve the training results we therefore sliced all images to 640x640 pixels beforehand. The sliced images were additionally modified by distorting all non-labeled regions of the slice by simply swapping pixel positions around randomly. This is done to prevent the model from seeing unlabeled data during the training process, as this would lead to a bad training performance. The weak grain detector is trained for

a specific task: distinguishing grains from other objects in the image. We finetuned the YOLO11 model *yolo11x* with 100 epochs of training using one class and all other parameters in default mode.

As already shown in Figure 10 the resulting bounding boxes of the first simple detector are still weak due to the limited amount of training data. In the following we use this simple detector to generate further training data. The procedure is always as follows: As the images have a very high resolution and the objects to be detected are relatively small, the YOLO detector with SAHI and a confidence threshold of 0.5 is used. This process produced numerous (weak) bounding boxes across the training images, which the SAM2 model sam2-hiera-large uses as input to find the outline of one grain per bounding box. The low confidence score of the YOLO11+SAHI detector sometimes results in many bounding boxes which may lead to faulty segmentations, as shown in Figure 12. The rectangles are the bounding boxes found by the detector. The dashed lines are corresponding outlines of the SAM2 model. Green colored lines indicate a higher score compared to the red colored detections. To compare the detections a mean score is calculated by $\overline{S} = 0.5 (S_{\text{YOLO}} + S_{\text{SAM2}})$ where S_{YOLO} is the resulting confidence score of the YOLO11+SAHI detector and S_{SAM2} the score of the SAM2 detection respectively. To find the faulty detections, we calculate the Intersection over Minimum (IoM) for all pairs of detections A and B with

$IoM = \frac{\operatorname{area}(A \cup B)}{\min(\operatorname{area}(A), \operatorname{area}(B))}.$

If for a pair of contours IoM > 0.3 is true, the contour with the lower score \overline{S} is discarded (shown in red in Figure 12).



Figure 12: Example of postprocessing contours. Rectangles: detected grains. Dotted curves: contours found by SAM2. Green: kept. Red: discarded.

The new bounding boxes are defined by the extend of the contours. This procedure of boosting the amount of training samples results in a huge amount of new training samples. In order to be able to assess the performance, two different experimental setups were carried out.

Training Setup 1. Only the training data from image acquisition session 1 is used. These are 44 images from 22 classes. Boosting leads to approx. 25000 new training labels. The *yolo11x* model is fine-tuned again with 100 epochs of training, but now including all 22 classes for detection.

Training Setup 2. In addition to the 44 images from 22 classes from acquisition session 1, the training data from acquisition session 2 is now also used. This is an additional 45 images from 23 classes. The additional class (the colored tube segments) was also automatically labeled with the simple grain detector. This worked well and emphasizes the good performance of the stack of YOLO and SAM2.

3.3 Results & Outlook

For the test of the model obtained from *training setup 1* all of the 44 test images of image acquisition session 1 are used as input and processed as follows:

- detect bounding boxes with the fully trained 22 class YOLO11+SAHI detector
- find segmentations inside of each bounding box with the SAM2 model
- postprocess detections to discard bad detections (cf. section 3.2)

This procedure yields exactly 24692 detected contours of which 24691 are correctly classified, i.e. a test accuracy of 99.996%. As shown in Figure 13, the results are very accurate.

These are really good results, but could be influenced by the uniform arrangement in session 1. Only images from a single shooting situation were used, which might have made recognition an easy task.

For training setup 2 two different tests were conducted following the same procedure as before: Here, the test data from image acquisition sessions 1 and 2 were used. This resulted in exactly 47679 detected contours, of which 100% are identified as the correct class. This is another outstanding result, which could again be attributed to the similarity of the test data to the training data. To counteract this, another test run was carried out with the data from recording session 3. As well as in the previous test results, no data from this session was ever used for training. But beyond that, the shots from this session represent a configuration of focus, aperture and exact camera position not previously used in any training situation. For this test images the upper as well as the lower part of the sample cup are used. The test on this data leads to exactly



Figure 13: Cuttings from images of the 16 fertilizers used in the project.

47736 contours of which 47693 were correctly recognized yielding a recognition rate of 99.901%. Provided the photo box presented in this paper is used in similar situations, this result can be considered reliable for a real-world application too.

Future work will now involve further analysis of the resulting contours in terms of size, shape and statistical distribution of grain sizes.

4 CONCLUSIONS

We presented a CV-ML pipeline composed of stateof-the-art tools, namely YOLO11 and SAM2, for the detection and classification of fertilizer grains. Using a Raspberry Pi HQ Camera mounted in a purposebuilt imaging box with a simple diffuse light source, the system setup is fairly simple. Training the detector and classifier in an active learning approach from only a few manually labeled images resulted in a test accuracy of 99.996%, despite the simplicity of our proposed setup. Although our focus was the classification of fertilizer as part of a bigger system, our proposed setup and pipeline can also serve as a template for other similar applications.

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