# SEGMENTATION OF SES FOR PROTEIN STRUCTURE ANALYSIS 

Virginio Cantoni, Riccardo Gatti and Luca Lombardi<br>University of Pavia, dept. of Computer Engineering and Systems Science,Via Ferrata 1, Pavia, Italy

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#### Abstract

The morphological complementarities of molecular surfaces provides insights for the identification and evaluation of binding sites. A quantitative characterization of these sites is an initial step towards protein based drug design. The final goal of the activity here presented is to provide a method that allows the identification of sites of possible protein-protein and protein-ligand interaction on the basis of the geometrical and topological structure of protein surfaces. The goal is to discover complementary regions (that is with concave and convex segments that match each others) among different molecules. In particular, we are considering the first step of this process: the segmentation of the protein surface in protuberances and cavities through an approach based on an analysis of the molecule Convex Hull and on the Distance Transform.


## 1 INTRODUCTION

An important research activity, with the large set of proteins in the current Protein Data Bank (PDB), is the prediction of interactions of these molecules by the discovery of similar or of complementary regions on their surfaces. When a novel protein with unknown functionalities is discovered, bioinformatics tools are used to screen huge datasets of proteins searching for candidates binding sites. More specifically, if a surface region of the novel protein is similar to that of the active site of another protein with known function, the function of the former protein can be inferred and also its molecular interaction can be predicted. Active sites are generally in concave and deep spots of the surface that are called "pockets".

Much work has been done on the identification and the analysis of the binding sites of proteins using various approaches based on different protein surface descriptions and matching strategies. The techniques employed are ranging from geometric hashing of triangles of points and their associated physico-chemical properties (Shulman, 2004), to clustering based on a representation of surfaces in terms of spherical harmonic coefficients (Glaser, 2006) or by a collection of spin-images (Bock, 2007 - Bock, 2008) or by context shapes (Frome, 2004), to clique detection on the vertices of the triangulated
solvent-accessible-surface (SAS) (Akatsu, 1996), to local surface 'buriedness' evaluation (Brady, 2000).

The goal of this work is to segment the protein surface in protuberances and cavities. This segmentation is based on the Distance Transform (DT) applied to the volume obtained subtracting the molecule to the its Convex Hull (CH). Once obtained protrusions and inlets, for each segment, a few features are provided including area and volume of the inlet, area and circumference of the pocked mouth opening, curvature (Cantoni, 2009) and travel depth. These features are the basic parameters for the first screening of compatible sites. A more precise subsequent experimental analysis on a limited subset of cases must be then applied.

This paper is organized as follows: section two shows a survey of approaches for segmentation and analysis of protein surfaces through the convex hull; in section three is introduced the solution proposed, then in section four a few results on artificial testimages and on true proteins molecules are presented. The final section, section five, provides a few concluding remarks and briefly describes our planned activity in the near future.

## 2 SURFACE ANALYSIS SEGMENTATION THROUGH THE CONVEX HULL

The CH of a molecule is the smallest convex polyhedron that contains the molecule points. In $\mathbf{R}^{3}$ the CH is constituted by a set of facets, that are triangles, and a set of ridges (boundary elements) that are edges. A practical $O(n \log n)$ algorithm for general dimensions CH computing, is Quickhull (Barber, 1996), that uses less space and executes faster than most of the other algorithms.

The CH approach for molecular segmentation is not new. The first paper applying this method, to authors' knowledge, is (Meier, 1995). The Quickhull algorithm, is applied to the SAS which is defined by the center of a water-sized probe sphere (usually with radius values ranging between 1.4 and $1.8 \AA$ ) 'rolling over the van der Waals surface of the atom'. The technique is based on two specificities: i) the tips lie directly on the CH surface: they are the common points between molecule and CH surfaces; ii) inlets and holes are 'normally' covered by large facets of the CH surface. Both specificities are not necessary conditions (the second is even not sufficient) to establish the existence of true tips and inlets: it is just a reasonable first hypothesis. Moreover, the technique for tip segmentation is based on a heuristic approach: each tip is extended on the outside facet for a distance determined by a global parameter.

Two other different approaches based on the CH of the atoms centers have been proposed by Edelsbrunner (Edelsbrummer, 1998) and Xie (Xie, 2007). Both apply the Dealunay triangulation technique, that in 3D have complex counterparts (3D tetrahedrons), to evaluate quantitatively some parameters.

The former, through a dual complex (alpha shape) analysis, provides a quantitative description of the microenvironments for protein structure based design. In particular, volume and area of pockets, area and circumferences of mouth opening, are evaluated. For the pockets identification, it is used the discrete flow method, that is the presence of Delaunay's tetrahedra disjointed to the dual complex. In particular, for segmentation purposes, some geometrical and topological rules allow the discrimination between two neighboring tetrahedra, satisfying the previous constraint. Note that, not all the inlets are identified as pocket (the one for which the discrete-flow pours to the outside of the CH ).
The latter approach is based on a simplified description that requires only the $\mathrm{C} \alpha$ atoms to
represent the protein structure in order to speed-up the computation (making the new representation "scalable to a large data set ... yet robust enough to handle the intrinsic properties of protein flexibility"). Moreover, the notion of geometric potential is introduced: this figure quantitatively describes the microenvironment on the basis of two heuristic parameters and allows a fast and effective discrimination for active sites.


Figure 1: Common 2D representations of surface models for protein's molecules: i) in green the van der Waals surface, directly produced from the atom's locations through the van der Waals radii; ii) in red the SolventExcluded Surface SES (also known as the molecular surface or Connolly surface) generated by the envelope of a rolling sphere over the van der Waals surface (The radius of the solvent sphere is usually set to the approximate radius of a water molecule having a van der Waals radius of $1.4 \AA$ ); iii) in blue the solvent accessible surface (sometimes called the Lee-Richards molecular surface) generated by the center of the solvent sphere rolling over the van der Waals surface; iv) in brown the convex hull, that coincides also with the SES having a sphere with an infinite radius.

The CH is also the reference surface for the molecules analysis based on the 'travel depth'. The travel depth parameter (Coleman, 2006 - Giard, 2008), with reference to the SES (see figure 1), is defined as the shortest path accessible for a solvent molecule between the protein convex hull and a given point that belongs to the 'active' region of interest (ROI) delimited by CH and SES. It represents the physical distance that a 'sufficiently' small molecule has to travel to approach a surface position (the pockets bottom are usually the points of interest). It is particular the case of tunnels, i.e. when pockets have no 'bottom', in which the molecule can travel through the entire protein and the travel depth is delimited by two points belonging to the CH. In particular (Coleman, 2006) introduced a technique for computing the travel depth on the basis of a peculiar distance transform implementation in the ROI defined above. The
implementation proposed by Giard et al. has the goal of speed-up the travel depth computation through a surface-based propagation algorithm that should be in general faster than the volume-based DT.

## 3 TUNNELS AND POCKETS DETECTION

In the discrete space the protein and the CH are defined in a cubic grid V of dimension L x $\mathrm{M} \times \mathrm{N}$ voxels. Note that the grid is extended one voxel beyond the minimum and maximum coordinate of the SES in each orthogonal direction (in this way both SES and CH borders are inside the V border). The voxel resolution adopted is $0.25 \AA$, so as to be small enough to ensure that, with the used radii in biomolecules atoms, any concave depression or convex protrusion is represented by at least one voxel.

Let us call R the region between the CH and the SES (the concavity volume (Borgefors, 1996)), that is:

$$
\begin{equation*}
\mathrm{R}=\mathrm{CH} \cap \overline{\mathrm{SES}} \tag{1}
\end{equation*}
$$

Let us call $\mathrm{B}_{\mathrm{CH}}$ the set border voxels of CH , that is:

$$
\begin{equation*}
\mathrm{B}_{\mathrm{CH}}=\mathrm{CH}-[\mathrm{CH} \square \mathrm{~K}] \tag{2}
\end{equation*}
$$

Where■ represent the erosion operator of mathematical morphology and K the discrete unitarian sphere (in the discrete space a $3 \times 3 \times 3$ cube!).

Within the region R the following propagation is applied:

$$
\begin{aligned}
& \mathrm{D}_{i}=\left\{\begin{array}{l}
1 \text { iff } \mathrm{i} \in \mathrm{~B}_{\mathrm{CH}} \\
0 \text { otherwise }
\end{array}\right\} \\
& \mathrm{A}=\mathrm{B}_{\mathrm{CH}} ; \\
& \mathrm{N}=(\mathrm{A} \oplus \mathrm{~K}) \cap \mathrm{R} ; \\
& \mathrm{E}=\mathrm{N}-\mathrm{A} ; \\
& \text { while } \mathrm{E} \neq \varnothing \text { do } \\
& \forall \mathrm{e} \in \mathrm{E}: d_{e}=\min _{n \in n n_{e}}\left(d_{n}+w_{n}\right) ; \\
& \mathrm{A}=\mathrm{N} ; \\
& \mathrm{N}=(\mathrm{A} \oplus \mathrm{~K}) \cap \mathrm{R} ; \\
& \mathrm{E}=\mathrm{N}-\mathrm{A} ; \\
& \text { done }
\end{aligned}
$$

where:
i. A represents the increasing set of voxels contained in R ;
ii. E corresponds to the recruited set of near neighbors of A contained in R (i.e. the voxels reached by the last propagation step);
iii. $\min _{n \in n n_{e}}\left(d_{n}+w_{n}\right)$ represents the minimum value among the distances $d_{e}$ in the near neighbors belonging to D already
defined, incremented by the displacement $w_{j}$ between the locations ( $\mathrm{e}, \mathrm{n}$ ): that is, if e and n have a common face $w_{n}=1$; if e and n have a common edge $w_{n}=\sqrt{2}$; if e and n have a common vertex $w_{n}=\sqrt{3}$. In three dimensions, the total number of the near neighbor elements of p is 26: six of them that share one face and have distance equal to 1 from the voxel p , twelve neighbors that share only an edge and are at distance $\sqrt{2}$, and eight that share only a vertex and are at distance $\sqrt{3}$ always from voxel $p$. At each iteration, new voxels, inside $R$, are reached by the propagation process and the value they take is determined by the neighbor distance (from the convex hull) and the voxels distance from the neighbor involved; this in order to simulate an isotropic propagation process and the proper distance evaluation.
iv. $\quad \mathrm{E}=\varnothing$ corresponds to the regime condition: no other changes are given and the connected component of R , adjacent to the border $\mathrm{B}_{\mathrm{CH}}$, is completely covered.

The values in D represent the distance of each voxel of A from the border of $\mathrm{B}_{\mathrm{CH}}$ and A corresponds to the connected component of R adjacent to the border.
Having A, it is possible to easily identify and eventually remove the cavities $C$, that are the volumes completely enclosed in the macromolecule M:

$$
\begin{equation*}
\mathrm{C}=\mathrm{CH}-\mathrm{A}-\mathrm{M} \tag{3}
\end{equation*}
$$



Figure 2a: A 2D example for tunnels and pockets detection composed of three connected components (in brown). The closed curve in black corresponds to the convex hull, and the border BCH in gray embodies the area under analysis and is the starting set of voxels for the propagation process.


Figure 2b: Results achieved after the propagation phase. Three pockets are identified, in blue, red and violet respectively. For the other three sides the propagation process converges: firstly there is the merging of the green and yellow waves, then the yellow and brown waves, and the complete coverage of the accessible areas is achieved in location L and I respectively. Note the three internal inaccessible components, in white, with labels $\alpha, \beta, \gamma$.


Figure 2c: Final segmentation, showing the tunnel (in green) and the three detected pockets.


Figure 2d: Another representation of the results after the propagation phase. The letters B, D, F correspond to the top of three pockets (see also figure 2 b )). The letters A, C, and E correspond to three local tops that are adjacent to important inlets. The letters $\mathbf{g}$ and $\mathbf{h}$ corresponds to convergences towards the tops I and L which identify a threelobate tunnel.

In order to separate the different pockets and tunnels the volume A must be partitioned into a set of disjoint segments $\mathrm{P}_{\mathrm{SES}}=\left\{\mathrm{P}_{1}, \ldots, \mathrm{P}_{\mathrm{j}}, \ldots, \mathrm{P}_{\mathrm{N}}\right\}$, where N is the number of inlets. The partition must satisfy the following condition:

$$
\begin{gather*}
P_{i} \cap P_{j}=\varnothing, i \neq j  \tag{4}\\
P_{1} \cup \cdots \cup P_{j} \cup \cdots \cup P_{N}=A \tag{5}
\end{gather*}
$$

As can be easily extended form the 2D example of Figure 2, starting from the total set of convex hull facets, several waves are generated and propagation proceeds up to the complete coverage of the volume A: the connected component of R adjacent to the border. During the propagation phase two sets of salient points are identified: local tops LT (represented by capital letters in Figure 2) and wave convergence WC points (lower case).

The LT set is exploited for the segmentation process. The cardinality of LT corresponds to $\mathrm{N}_{\max }$ the maximum number of segments/inlets that can be considered. The effective number of segments, that determines obviously the number and the morphology of pockets and tunnels, is found out on the basis of two heuristic parameters: i) the minimum travel depth value of the local tops $\mathrm{TD}_{\mathrm{LT}}$; ii) an evaluation of near neighbor pivoting effects PEs. The threshold $\mathrm{TD}_{\mathrm{LT}}$ is introduced because the surface's irregularities and the digitalization process produce small irrelevant spurious cavities. The thresholds PEs take into account morphological aspects insight important cavities and can be characterized by two different features: the nearness of others, more significant, local tops ( $\tau_{1}$ ) and the
relative values of the local-top travel-distance $\left(\tau_{2}\right)$. in general faster than the volume-based DT.

## 4 IMPLEMENTATION AND RESULTS

We start from the 'space-filling' representation of the protein, where atoms are represented as spheres with their van der Waals radii (this representation is directly derived from PDB files which supply the ordered sequence of 3D positions of each atom's center). Figure 3 shows the image produced by our package for Apostreptavidin Wildtype CoreStreptavidin with Biotin structure (1MK5 in PDB).


Figure 3: On the left the 'space filling' representation of 1MK5. The colors follow the standard CPK scheme. On the right the correspondent secondary structure representation.

A first critical decision is the space resolution level for the analysis. The results presented here are given with a resolution of $0.25 \AA$, which entails a van der Waals radius of more than five voxels to the smallest represented atoms. The algorithm is then applied to the SES obtained from the quoted surface, after the execution of a closure operator using a sphere with radius of $1.4 \AA$, about 6 voxels, (corresponding to the conventional size of a water molecule) as structural element. It is worth to point out that this closure operation excludes possible passage through apertures with a section of less than $6.15 \AA^{2}$ (about 99 voxels). Note that, as it has been mentioned in section 3, the grid is extended one voxel beyond the minimum and maximum coordinate of the SES in each orthogonal direction (in this way both SES and CH borders are inside the V border).

Two other parameters characterize the execution: the minimum passage section $\theta_{1}$ (obviously $\theta_{1} \geq 100$ ); the maximum mouth aperture $\theta_{2}$. The former has been fixed on a heuristic base to 150 Voxels (which for a circle corresponds to a radius of 7 voxels). The latter has been applied with two different values: 2000, 7500 .


Figure 4: A representation of the results after two segmentation phases with $\theta_{2}$ equal to 2000 and 7500 in a) and b) respectively. Note that no tunnel is present, and 9 pockets have been identified and differently colored. In particular the first three pockets in the top figure are then represented in figures 5, 6 and 7 corresponding to $\theta_{2}=2000$ and the first two pockets in figures 8 and 9 having $\theta_{2}=7500$.

In Figure 4 the results of the segmentation process are given. This process is executed in two phase: in the onward propagation the set of the pocket's local top is identified; later a backward parallel propagation from each of the tops with identify all the pockets is executed. Finally, it is applied the near neighbor pivoting process, which (with $\tau_{1}=10$ and $\tau_{2}=5$ ) leads to the final result shown. Figures 5-9 show the pockets with the highest travel distance (obviously the parameters of reference can be one of the others features - e.g. the pocket volume -, or even a combination of features e.g. travel distance and pocket volume-) for different values of the parameter $\theta_{2}$. Each pocket is represented with a lateral and frontal view (respectively on the left and on the right side of each figure).
As it can seen from figure 4 , having $\theta_{1}=150$ voxels no tunnels are present, but there are several well characterized pockets which can be easily characterized and evaluated. In particular the dependence of the results of the segmentation process from the different parameters is pointed out. The algorithms has been tested on a $2.20 \mathrm{GHz} \times 86$ Intel processor the computing time is about 12 seconds for the protein 1 MK 5 with 127 residues and 1844 atoms.


Figure 5: The pocket with the highest travel depth (highest distant from the CH ): 57 , with the control parameter $\theta_{2}$ to the value: 2000 . Note that the red component testifies that the 'pocket mouth' is close to the CH .


Figure 6: The pocket with the second highest travel depth: 35 , with $\theta_{2}=2000$. Note that also in this case, the 'pocket mouth' is close to the CH.


Figure 7: The pocket with the third highest travel depth: 34, with $\theta_{2}=2000$. Note the absence of the red component: the 'pocket mouth' is at a distance of 20 voxels from the CH .


Figure 8: The pocket with the highest travel depth achieved with $\theta_{2}=7500$. Note that for this value of the control parameter the two highest pocket achieved with $\theta_{2}$ $=2000$ are fused together realizing a large 'pocket mouth' more close to the CH .


Figure 9: The pocket with the second highest travel depth with $\theta_{2}=7500$. Note that this is the evolution of the previous third pocket and is characterized by a large 'pocket mouth' and the presence of the red component (that is the mouth is closer to the CH ).

## 5 CONCLUSIONS

The final goal of the activity here presented is to provide a method that allows the identification of sites of possible protein-protein and protein-ligand interaction on the basis of the geometrical and topological structure of protein surfaces. The goal is then to discover complementary regions (that is with concave and convex segments that match each others) among different proteins. In particular, we are considering the first step of this process: the segmentation of the protein surface in various pockets and tunnels. The next step of our activity is related to the characterization of each of the extracted segment through morphological and topological quantitative descriptors (including travel depth, mouth aperture, curvature, volume) that can be combined with the local biochemical features (types of residues and their characteristics) to detect and specialize the active sites of a protein.

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